

## Novel measurements of atmospheric iodine species by resonance fluorescence

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**Abstract** A field instrument has been developed for the purpose of measuring gas-phase atmospheric iodine species in the marine boundary layer. Vacuum UV resonance-fluorescence (RF), generated using a microwave discharge lamp, is employed to detect atomic iodine via the  $(5p^46s)-(5p^5)$  transitions around 178–184 nm. The system can be operated in two modes; either to directly measure ambient iodine atoms, or to measure the total photolabile iodine loading of ambient air, through broadband visible photolysis of photolabile iodine-containing species, with subsequent RF detection of the iodine atoms released. In both cases the instrument allows for the in situ measurement of the species detected, which is advantageous for gathering information concerning their local sources and distribution. The instrument is calibrated through generation of a known concentration of iodine atoms from the photolysis of  $I_2$  using a mercury pen-ray lamp. The instrument was deployed for the first time in August 2007 at Mace Head on the west coast of Ireland; initial results from this field trial are presented. Ambient iodine atoms were measured at levels up to  $22 \pm 4.8$  ppt during the day, coinciding with the lowest tides, when *Laminaria* seaweed beds were exposed. The total photolabile iodine loading was also measured during several night-time and day-time periods and was found to correlate inversely with tidal height. Inferred  $I_2$  concentrations based on these measurements indicate levels of several hundred ppt at the Mace Head site. These measurements represent the first direct observations of ambient iodine atoms and measurement of total photolabile iodine in the atmosphere.

**Keywords** Iodine · Iodine monoxide · Mace head · Marine boundary layer · Resonance-fluorescence

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## 1 Introduction

The potential role of iodine species in the atmosphere has been considered for a number of years (e.g. Chameides and Davis 1980; Solomon et al. 1994). More recently, iodine species have been shown to affect the chemistry of the coastal marine boundary layer (MBL, e.g. Platt and Hönninger 2003; Carpenter 2003; von Glasow and Crutzen 2007), and have been measured in the polar boundary layer (Saiz-Lopez et al. 2007). In the marine environment, molecular iodine and alkyl iodides such as  $\text{CH}_3\text{I}$  and  $\text{CH}_2\text{I}_2$  are emitted by micro and macro algae and undergo photolysis on timescales of seconds ( $\text{I}_2$ ) to days ( $\text{CH}_3\text{I}$ ) releasing their constituent iodine atoms, which then react with ozone, forming the iodine monoxide radical (IO).



where  $\text{R}=\text{CH}_3, \text{CH}_2\text{I}, \text{C}_2\text{H}_5$  etc.



IO is known to react with itself,  $\text{HO}_2$  and  $\text{NO}_2$  in cycles which may catalytically destroy tropospheric ozone (Chameides and Davis 1980; Vogt et al. 1999). As a result of such reactions, the  $\text{OH}:\text{HO}_2$  and  $\text{NO}:\text{NO}_2$  ratios may also be altered, which will in turn affect the oxidative capacity of the atmosphere (Bloss et al. 2005). It has also been suggested that the IO radical can polymerise via  $\text{OIO}/\text{I}_2\text{O}_2$  to form larger condensable  $\text{I}_x\text{O}_y$  molecules, leading to aerosol growth and/or new particle formation, with potential implications for MBL cloud condensation nuclei abundance and hence radiative balance (O'Dowd et al. 2002).

Field measurements of species such as  $\text{I}_2$ , IO, and the alkyl iodides ( $\text{CH}_3\text{I}$ ,  $\text{CH}_2\text{I}_2$  etc.), have been made previously in the marine boundary layer:  $\text{I}_2$  has been measured using long-pass differential optical absorption spectroscopy (DOAS; Saiz-Lopez and Plane 2004; Peters et al. 2005); however, this technique is associated with significant spatial and temporal averaging. IO and OIO have also been measured at various coastal sites using DOAS (e.g. Alicke et al. 1999; Saiz-Lopez et al. 2004). An issue for interpretation of DOAS observations, particularly at coastal sites such as Mace Head, has been the spatial distribution of iodine species: the DOAS beam path (usually several kilometres in length) may encompass both the intertidal zone and the open ocean, which can lead to ambiguity in the identification of the source of the iodine species (O'Dowd and Hoffmann 2005). Recently, new point measurement/in situ methods have been developed to measure OIO and  $\text{I}_2$ , via broadband cavity ringdown spectroscopy (Bitter et al. 2005) and IO, via laser-induced fluorescence (Whalley et al. 2007), however there has, as yet, been no direct (or indirect) atmospheric observations of atomic iodine, or the total photolabile iodine content.

The technique of resonance fluorescence (RF) for the detection of atomic iodine has been widely applied in laboratory experiments (e.g. Plane and Husain 1986; Ingham et al. 2000; Gross et al. 2000), but has not previously been applied in the form of a field measurement instrument. For laboratory experiments, achieving a high sensitivity, and therefore, low limit of detection, may not be as critical as for field measurements, and the concentrations sampled are in general several orders of magnitude larger ( $10^9$ – $10^{12}$  cf.  $10^7$ – $10^8$   $\text{atom cm}^{-3}$ ). Typically the limit of detection for laboratory instruments is of the order  $10^9$ – $10^{10}$   $\text{atom cm}^{-3}$  or higher, raising challenges for field measurements; however the RF technique exhibits several advantages that make it an excellent choice for application as a

field instrument: high sensitivity, selectivity, low power consumption and potential for the components to be made lightweight and portable.

Atomic resonance fluorescence has been utilized for the detection of the other halogen species in the atmosphere, particularly the polar stratosphere. Anderson and co-workers developed a balloon-borne resonance-fluorescence instrument for the detection of ClO (by titration with NO and subsequent RF detection of the Cl atoms) in the stratosphere (Anderson et al. 1977, 1980). A similar instrument was also developed for the ER-2 aircraft to monitor stratospheric chlorine species (Brune et al. 1988, 1989a), with limits of detection better than 1 pptv. Subsequent developments allowed the detection of Cl<sub>2</sub>O<sub>2</sub>, via thermal decomposition followed by ClO titration with NO (Stimpfle et al. 2004). The chemical-conversion resonance-fluorescence (CCRF) technique has also been used for measurement of stratospheric BrO (Brune et al. 1989b) and more recently has been employed to detect ClO, Cl<sub>2</sub>O<sub>2</sub> and BrO on the M55-Geophysica (von Hobe et al. 2005, 2006).

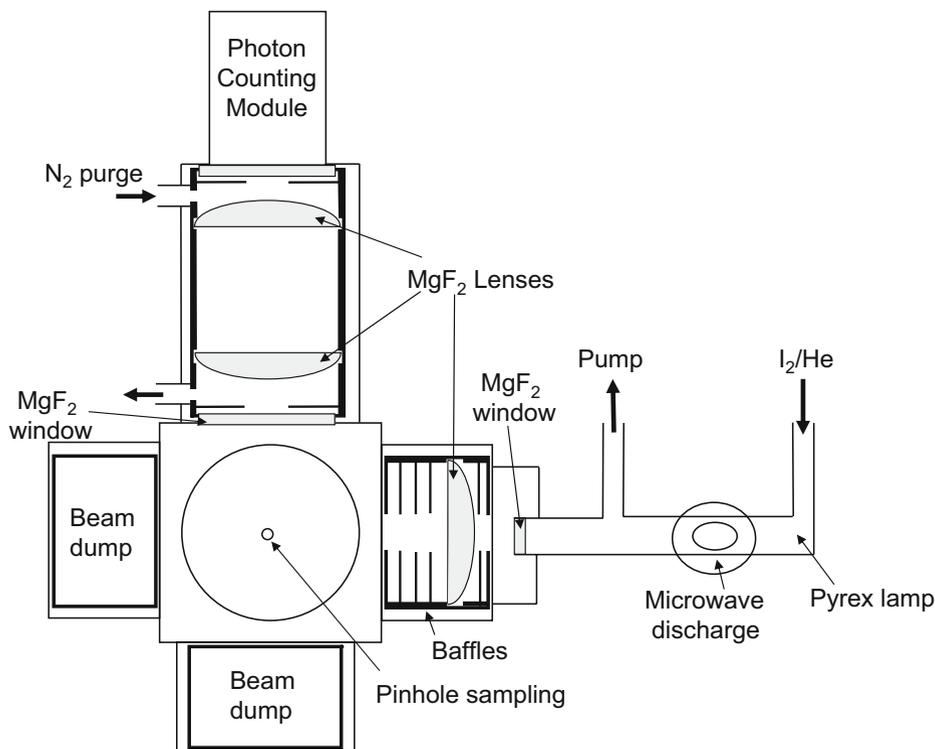
## 2 Instrument

The resonance lines of atomic iodine fall in the vacuum ultraviolet wavelengths in the range 170 – 190 nm. The two most important transitions are the (5p<sup>4</sup> 6s) <sup>2</sup>P<sub>3/2</sub>–(5p<sup>5</sup>) <sup>2</sup>P<sub>3/2</sub> at λ=178.3 nm, and the spin-forbidden (and, therefore, 15–20 times weaker) (5p<sup>4</sup> 6s) <sup>4</sup>P<sub>5/2</sub>–(5p<sup>5</sup>) <sup>2</sup>P<sub>3/2</sub> transition at λ=183.0 nm (Brewer and Tellinghuisen 1971). Absorption by molecular iodine, I<sub>2</sub>, of light in the region between 180 and 200 nm through the highly structured (D←X) system will lead to population of the I<sub>2</sub> D <sup>1</sup>Σ<sub>u</sub><sup>+</sup> state which fluoresces in the range 200–500 nm (O'Grady et al. 1982). As the photomultiplier used in this instrument detects radiation up to around 320 nm, a proportion of any I<sub>2</sub> fluorescence will be detected; any I<sub>2</sub> present therefore leads to an interference in the I-atom fluorescence signal but, as will be described later, this can be taken into account during calibration, and the sensitivity towards I<sub>2</sub> may be used advantageously. The instrument response to I<sub>2</sub> is, however, typically three orders of magnitude lower than that towards atomic iodine.

The instrument comprises a microwave discharge lamp and collimating optics to supply the excitation radiation, a fluorescence cell, detection optics, a photon counting module, data acquisition system and a separate calibration unit. Figure 1 shows a schematic diagram of the fluorescence cell setup.

### 2.1 Excitation radiation

Continuous-wave excitation radiation was generated in a Pyrex lamp using a microwave discharge to dissociate I<sub>2</sub> and produce excited I atoms. A small flow (10 sccm) of high purity helium (CP grade, >99.999% purity, BOC) regulated with a mass flow controller was continuously drawn through the lamp by a rotary vane vacuum pump. A small amount of I<sub>2</sub> vapour is entrained into the helium flow via a needle valve from a finger trap containing a few crystals of iodine. The pressure in the lamp was held at around 1 Torr and monitored with a 10 Torr capacitance manometer. The lamp is sealed with an MgF<sub>2</sub> window to allow transmission of the vacuum-UV radiation. The microwave discharge plasma is produced using an Evenson-type cavity supplied with 60 W of power (EMS Microtron 200). The lamp was kept clean from impurities by filling to above ambient pressure with helium when not in use. Periodically the lamp was washed with water to remove build-up of iodine on the inside surfaces, particularly on the exit window. Once the radiation has exited the lamp it was collimated using an MgF<sub>2</sub> plano-convex lens (Crystran Ltd.) and several baffles



**Fig. 1** Schematic diagram (plan view) of the RF instrument for the detection of atomic iodine. The main cell is constructed from a matt black anodised aluminium block. MgF<sub>2</sub> windows and lenses are used to allow transmission of the vacuum ultraviolet light. The pinhole is conically shaped with an aperture diameter of 0.8 mm. The lower face of the cell (not shown) houses connections to the pump, throttle valve and pressure gauge

(aperture diameter of 5 mm) before entering the fluorescence cell. Beam dumps were mounted within the cell opposite, and orthogonal to, the resonance lamp, to absorb scattered light and reduce the background signal (hence improving the limit of detection).

## 2.2 Fluorescence cell

The cell itself is an aluminium cube (65×65×65 mm), with 25 mm diameter holes bored through each face, anodised matt black to reduce scattering of light from the walls. The air sample is drawn through the cell at a rate of 5.6 slm through a conical pinhole (aperture diameter=0.8 mm), using a throttled rotary vane vacuum pump. Experimental tests revealed that the lowest cell pressure achievable with this setup (around 20 Torr using a 0.8 mm pinhole) gave the optimum sensitivity towards iodine atoms. Collisional quenching will increase with cell pressure, but will be balanced by increased total number density (as there is no time-resolved aspect to the fluorescence collection in this system). At significantly higher pressures, the increased transit time experienced by the atoms may allow chemical losses to become significant. The presence of a higher number density of O<sub>2</sub> and other molecules will also lead to increased scattering of the excitation light (raising the limit of detection) and absorption of the fluorescence light. Different sized pinhole apertures were tested (0.4–0.8 mm), in order to attain lower pressures in the cell. However, the sensitivity

was found to be best with a 0.8 mm diameter nozzle—likely to be a consequence of the optimum overlap of the expansion of the sampled gas within the cell, the excitation radiation and the solid angle sampled by the detection system. The pressure in the cell was therefore held in the range 20–22 Torr, and was monitored with a capacitance manometer.

The collection optics axis was separated from the fluorescence cell with an  $\text{MgF}_2$  window, and was flushed with dry nitrogen to eliminate absorption of the light by  $\text{O}_2$  or water vapour. The fluorescence was collected with two  $\text{MgF}_2$  plano-convex lenses (focal length 58 mm at 180 nm, Crystran Ltd.) and focussed onto the photocathode of a photon counting module (MP921P, Perkin Elmer), which encompasses a channel photomultiplier (CPM), high voltage supply and amplifier/(factory set) discriminator/counting electronics. The module was powered by a regular laboratory power supply unit at 5 V. The CPM was set to count for 1 second time periods. Typical dark count levels were of the order of  $1 \text{ s}^{-1}$ .

### 2.3 Data acquisition

The signal collected at the CPM was recorded (in counts per second) continuously, and sent to a portable PC via the RS232 port. A USB-based DAQ module (USB-1208LS, Measurement Computing) was used to log auxiliary data every second, including cell pressure, lamp pressure, and microwave power. The flows through all the mass flow controllers were also logged. Various system temperatures (below) were monitored via thermocouples (type K) and a USB-based 8-channel thermocouple input module. A program to display and log all data was developed using the Delphi programming language. The instrument was housed in an enclosed lightweight aluminium frame (approximately  $80 \times 80 \times 120 \text{ cm}$ ), and the instrument was powered from a regular 13 A extension cable.

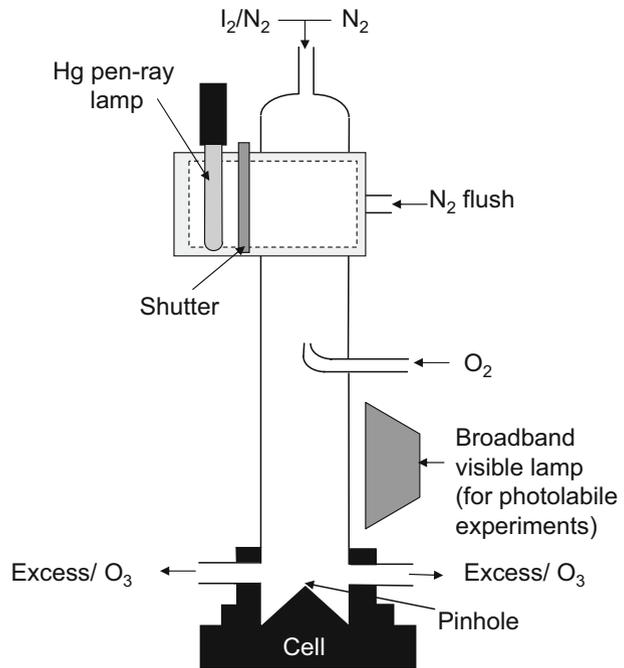
## 3 Calibration

Fluorescence detection is not an absolute technique and the instrument therefore requires calibration in order to relate the signal observed to a quantity of atoms or molecules present. This is achieved by sampling known concentrations of  $\text{I}_2$  or I and measuring the corresponding signal. The calibration setup, shown in Fig. 2, consists of a quartz flow-tube, gas supplies and mass flow controllers, and a mercury pen-ray lamp (LSP035, LOT Oriol). A fraction of the flow through the calibration tube was sampled into the RF cell, with the excess balance directed to an ozone analyser (during calibration) and vented.

### 3.1 Sensitivity to $\text{I}_2$

To calibrate the response of the instrument to  $\text{I}_2$  fluorescence, the signal is monitored upon addition of different amounts of  $\text{I}_2$  in a nitrogen bath gas to the calibration flow-tube. A controlled flow of nitrogen gas (10–50 sccm) was passed through a Pyrex U-tube packed alternately with  $\text{I}_2$  crystals (>99.5%, Aldrich) and glass beads in order to completely saturate the gas with  $\text{I}_2$  vapour. The tube was held in a water bath which acts as a thermal reservoir maintaining a stable temperature, and thus sustaining a constant vapour pressure from the  $\text{I}_2$  crystals. The temperature of the trap was monitored with a thermocouple (type K) and recorded using the USB-based thermocouple input module. The  $\text{I}_2/\text{N}_2$  gas mixture was diluted in a larger flow of  $\text{N}_2$  (10 slm) at the trap exit, and sent to the flow tube. The concentration of  $\text{I}_2$  in the flow tube may be determined using the vapour pressure of the  $\text{I}_2$  crystals at the recorded temperature (Antoine equation parameters taken from Stull 1947),

**Fig. 2** Schematic diagram of the calibration/photolysis flow tube

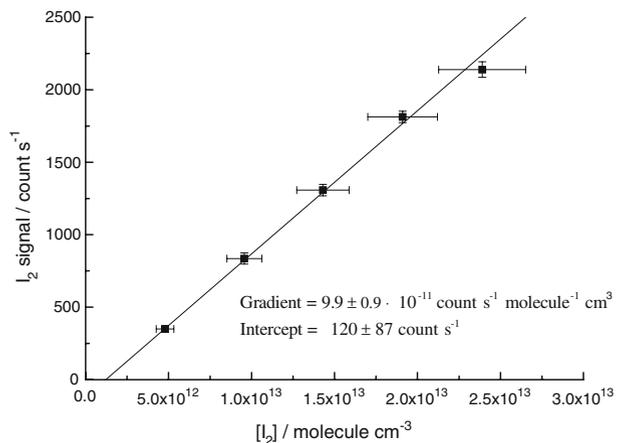


verified by UV absorption in a separate cell.  $I_2$  concentrations used in the calibration were in the range  $3\text{--}30 \times 10^{12}$  molecule  $\text{cm}^{-3}$ . A plot of  $I_2$  signal as a function of  $[I_2]$  can be seen in Fig. 3. During the field deployment the sensitivity to  $I_2$ ,  $C_{I_2}$ , was  $(8 \pm 0.8) \times 10^{-11}$  count  $\text{s}^{-1}$  molecule $^{-1}$   $\text{cm}^3$ .

### 3.2 Sensitivity to I atoms

The sensitivity of the instrument to atomic iodine was determined by photolysing  $I_2$  to generate known concentrations of atomic iodine. A mercury pen-ray lamp is used to photolyse  $I_2$  at 185 nm and 254 nm ( $\sigma_{185 \text{ nm}} = 1.93 \times 10^{-17}$   $\text{cm}^2$  per molecule;

**Fig. 3** Calibration plot showing measured  $I_2$  fluorescence signal as a function of  $[I_2]$  sampled from the calibration tube. The instrument sensitivity towards  $I_2$  is equal to the gradient of the line.  $y$ -Axis error bars are given as the standard deviation of the  $I_2$  fluorescence signal;  $x$ -axis error bars are given as the error on the determined concentration. The regression line fit is weighted to errors on both axes



$\sigma_{254\text{ nm}} = 1.13 \times 10^{-18} \text{ cm}^2$  per molecule; Saiz-Lopez et al. 2004); a quartz flow tube was used rather than Pyrex to allow transmission of the 185 nm light.

The pen-ray lamp was housed in an aluminium holder at a fixed distance from the fluorescence cell. The lamp was allowed to warm up to give a stable output, and was flushed with dry  $\text{N}_2$  to keep the lamp and housing at a stable temperature and prevent absorption of the 185 nm radiation by the oxygen in air. A shutter was mounted in front of the pen-ray lamp to allow modulation of the light output/photolysis of  $\text{I}_2$  without switching the lamp on and off. The background scattered signal was initially recorded; then,  $\text{I}_2$  was introduced into the calibration tube and the  $\text{I}_2$  fluorescence signal recorded. The shutter was then opened to expose the  $\text{I}_2$  flow to the pen-ray output, generating I atoms, and giving rise to a further increase in the observed signal. The concentration of iodine atoms generated in the calibration tube could be altered by changing the current supplied to the mercury pen-ray lamp, which in turn varies the flux of the lamp (unlike molecular oxygen, the cross section of  $\text{I}_2$  in this region is relatively smooth hence insensitive to small shifts in the lamp emission spectrum with current).

In order to determine the concentration of iodine atoms generated by photolysis of  $\text{I}_2$ , the flux from the pen-ray lamp at different applied currents must be known. The product of the lamp output flux at the wavelengths in question ( $F$ ) and the exposure time for which the calibration flow was within the illuminated region ( $t$ ) was determined using an ozone actinometry method similar to that described by Whalley et al. (2007): measurement of the ozone produced from photolysis of varying strength mixtures of  $\text{N}_2\text{O}$  in air, flowed through the calibration tube. The lamp flux was calibrated under the same flow conditions as were used for subsequent RF system calibrations, thus the exposure time was equivalent in both instances.

$\text{N}_2\text{O}$  was photolysed in the presence of  $\text{O}_2$  to generate ozone:



Some ozone will also be generated via photolysis of molecular oxygen at 185 nm:

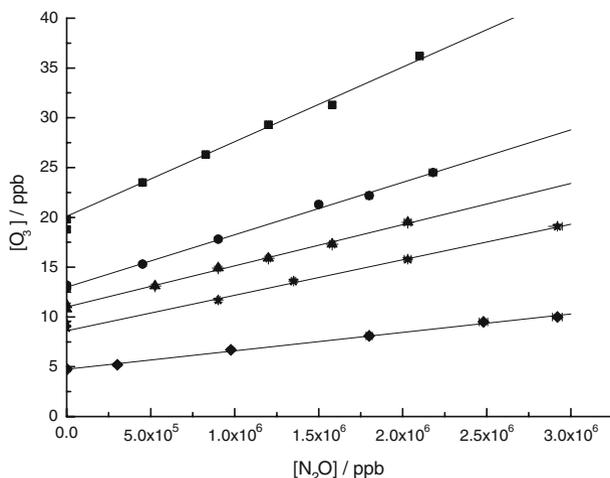


Gas from the flow tube was sent *via* the excess lines (Fig. 2) to a commercial ozone analyzer (Thermo Electron, TE49C or 2B Technologies, Model 205), with which  $[\text{O}_3]$  was monitored as the flow of  $\text{N}_2\text{O}$  was varied between 0–50 sccm. This procedure was repeated as a function of (measured) lamp current.

The product  $Ft$  may be calculated by

$$Ft = \frac{[\text{O}_3]_{\text{measured}}}{[\text{N}_2\text{O}]\sigma_{\text{N}_2\text{O}}} \quad (7)$$

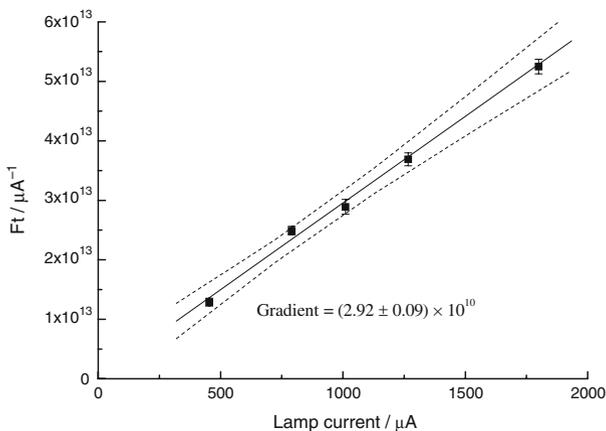
A plot of  $[\text{O}_3]$  as a function of  $[\text{N}_2\text{O}]$  for a range of lamp currents between 0 and 1500  $\mu\text{A}$  can be seen in Fig. 4; the product  $Ft$  can be calculated from the gradient divided



**Fig. 4** Determination of the 184.9 nm photolysis flux from the mercury pen-ray lamp at various supplied lamp currents using an actinometric method (Eq. 7). Plot shows  $[O_3]$  measured in the excess line as a function of  $[N_2O]$  in the calibration tube. A linear regression which takes into account errors in both  $x$ - and  $y$ -axes was applied to the data. Symbols represent different lamp currents ( $\mu A$ ): square=1,800; circle=1,267; triangle=1,010; star=790; diamond=454. Error bars are shown (but may be unclear because of their small magnitude)  $y$ -axis error bars represent uncertainty in the  $O_3$  measurement;  $x$ -axis error bars represent uncertainty in the flows and hence concentration of  $N_2O$ . The intercepts arise from the formation of ozone from the photolysis of  $O_2$

by the absorption cross-section of  $N_2O$  at 185 nm ( $\sigma_{185 \text{ nm}} = 1.43 \times 10^{-19} \text{ cm}^2$  per molecule). The absorption of  $N_2O$  at 254 is not significant ( $\sigma_{254 \text{ nm}} < 1 \times 10^{-23} \text{ cm}^2$  per molecule; Atkinson et al. 2004), and therefore this method determines only the flux of the 185 nm line from the lamp. The product  $Ft$  may then be plotted as a function of lamp current (see Fig. 5) to obtain a linear relationship between lamp current and lamp flux. The uncertainty associated with measurement of the 184.9 nm lamp flux is minimal, approximately 3%, as the uncertainties in  $[N_2O]$  and  $\sigma_{N_2O}$  are small.

**Fig. 5** Relationship between lamp photolysis flux and lamp current, derived from Fig. 4. The gradient of the line can be used to determine the lamp flux for a given lamp current. The  $y$ -error bars represent the uncertainty in the gradients shown in Fig. 4. Dashed lines indicate the 95% confidence limit on the linear regression



Once the flux emitted from the lamp is known, the concentration of iodine atoms formed upon photolysis of  $I_2$  can be calculated via

$$[I] = ([I_2] \times Ft \times \sigma_{(I_2, 185)} \times 2) + ([I_2] \times Ft \times \sigma_{(I_2, 254)} \times 2 \times 15) \quad (8)$$

where  $Ft$  is the product of the mercury lamp flux and the exposure time (at a given lamp current) and  $\sigma_{(I_2, 185)}$  and  $\sigma_{(I_2, 254)}$  are the absorption cross sections of  $I_2$  at 185 nm and 254 nm respectively. It is assumed that the quantum yield for photolysis of  $I_2$  is 2, which is consistent with quantum yield measurements below 340 nm (Atkinson et al. 2007). The factor of 15 accounts for the intensity of the 254 nm mercury line relative to that at 185 nm (Lide 2007). The ratio of  $I_2$  photolysed at 254 nm compared with that at 185 nm is 0.9:1.

The calibration factor for iodine atom detection, ( $C_I$  in units of  $\text{count s}^{-1} \text{atom}^{-1} \text{cm}^3$ ) may then be determined using

$$C_I = \frac{S_I}{[I]} \quad (9)$$

where  $S_I$ , the signal due to iodine atom resonance fluorescence, is given by

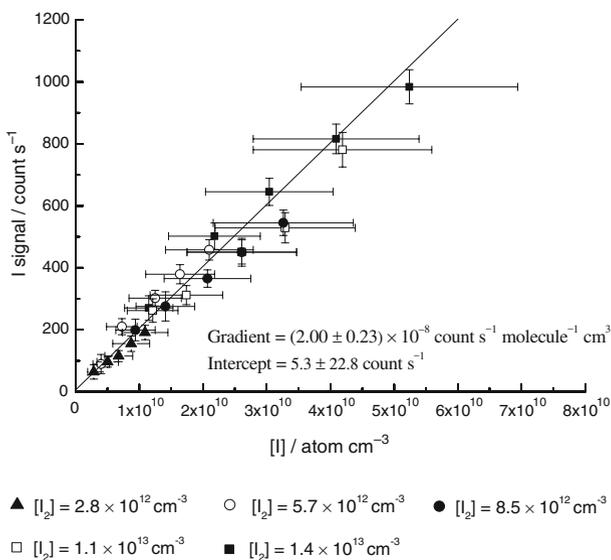
$$S_I = S_{\text{total}} - S_{I_2} - S_b \quad (10)$$

and  $[I]$  is determined as given in Eq. 7;  $S_{\text{Total}}$  is the signal when  $I_2$  is present and the shutter to the lamp is open,  $S_{I_2}$  is the signal from  $I_2$  fluorescence only and  $S_b$  is the background signal [comprising dark counts (typically  $<1 \text{ count s}^{-1}$ ), scattered light from the cell walls, air and aerosol, and any interference from other species]. Typically, five repeat points were measured at varying lamp currents and the calibration factor,  $C_I$ , is determined from the gradient of a plot of  $S_I$  vs.  $[I]$ .

The iodine atom calibrations were conducted in air, to best represent atmospheric conditions. However, it was not possible to use air as the bath gas in the flow tube, as any  $O_2$  passing in front of the mercury pen-ray lamp would be photolysed at 185 nm; the resulting O atoms and ozone would react with both  $I_2$  and I causing an interference in the calibration method. Instead,  $N_2$  was used as the bath gas for the  $I_2$  in the photolysis region, and  $O_2$  was added subsequently, through a separate injection port in the flow tube located after the photolysis region. The  $O_2$  flow was set to make up 21% (2.1 slm) of the total flow in the calibration tube with the remainder made up from  $N_2$  (7.9 slm). Experiments conducted in a bath gas of air showed a significant (approximately 30%) decrease in iodine atom sensitivity compared with experiments in nitrogen only, primarily as a result of the absorption of the excitation and fluorescence radiation by  $O_2$ .

The calibration conditions employed ( $I_2$  concentration and photolysis flux, and hence I atom concentrations) were necessarily limited by the sensitivity of the detection techniques available to determine the molecular iodine concentration (UV absorption spectroscopy) and the lamp flux (ozone actinometry—ozone monitored using a commercial UV-absorption monitor); consequently calibrations were performed at levels higher than the ambient concentrations of iodine atoms and molecular iodine encountered. To verify that the instrument response was constant and linear, and to ensure that quenching of the excited I atoms or absorption of the emitted fluorescence were not appreciably enhanced by the presence of  $I_2$ , calibrations were performed over as wide a range of  $I_2$  levels and photolysis flux values as the methodology and iodine vapour pressure would permit (see Fig. 6). These calibrations showed no significant change in sensitivity towards I atoms with  $[I_2]$  varied over the range  $2\text{--}20 \times 10^{12} \text{ molecule cm}^{-3}$ . Note that the error bars in the points shown in Fig. 6 arise from imprecision in the calibration factors (x-axis:  $I_2$  concentration, lamp flux,

**Fig. 6** Typical calibration plot showing I atom fluorescence signal as a function of [I] sampled from the calibration tube. The gradient of the line determines the instrument sensitivity towards I atoms. Symbols indicate calibration runs performed with various concentrations of I<sub>2</sub> in the flow tube. y-Axis error bars indicate the standard deviation of the I atom fluorescence signal; x-axis error bars indicate the uncertainty in the calculated concentration of [I], arising from the lamp flux measurement and the I<sub>2</sub> concentration determination. The regression line takes into account errors on both the x- and y-axes



flow rates) and the fluorescence signal (y-axis, including contribution from subtraction of I<sub>2</sub> fluorescence signal); the latter factor in particular is much larger under the calibration conditions than is the case in ambient air. The uncertainty in the regression line parameters reflects the fact that Fig. 6 is a composite of five separate calibration runs performed at different times with different I<sub>2</sub> concentrations, indicating the invariance of the instrument response. The uncertainty in the gradient of Fig. 6, 11.5%, is factored into the overall uncertainty in the reported I atom measurements.

The effect of water vapour on the sensitivity of the instrument was also investigated. The O<sub>2</sub> gas injected into the side arm was first bubbled through a trap containing water in order to saturate the gas; again it was necessary to inject the water vapour after the photolysis region to avoid any interference in the iodine chemistry from HO<sub>x</sub> radicals formed by water photolysis. H<sub>2</sub>O vapour was varied in the range 0–1.4% which is comparable to levels found in ambient air (during the measurements at Mace Head, reported below, the mean relative humidity was 88%, corresponding to a water vapour fraction of 1.4%). The instrument sensitivity was found to be constant (within uncertainty) over this range.

The Reynolds number which describes the flow in the calibration tube is calculated to be 650 at a total flow of 10 slm (in a tube of internal diameter 2.1 cm at a pressure of 760 Torr and a temperature of 298 K), and hence the flow is within the laminar regime. Under these conditions the axial velocity will be greatest at the centre of the calibration tube. In similar experimental arrangements (Aschmutat et al. 1994) this effect has been shown to lead to lower radical concentrations at the centre of the flow tube relative to the periphery (as the gas in the centre of the tube is exposed to the photolysis light for a shorter time). Because the lamp flux is measured by sampling ozone from the excess line (at the edge of the tube) this could lead to an overestimation of the lamp flux, and thus an underestimation of the calibration factor. A correction for this effect can be applied and may be termed the gas profile factor, *P*, equal to the ratio of the ozone concentration in the central region of the tube, drawn into the RF cell, to that in the excess flow measured by the ozone analyser.

However, tests to measure  $[\text{O}_3]$  at the centre of the calibration tube compared with at the edge of the tube, confirmed that in our experimental setup,  $P=1$ ; the injection of  $\text{O}_2$  from the side arm introduces sufficient turbulence to thoroughly mix the flow, effectively eliminating the concentration gradient. A consequence of this is that some wall-loss of iodine atoms, in the turbulence induced by the  $\text{O}_2$  addition, may occur, implying a potential underestimation of the system sensitivity—see below. During the field deployment at Mace Head, short calibrations were performed daily to monitor any significant change in the sensitivity of the instrument, with full calibrations as described above (data shown in Fig. 6) conducted in Leeds. The calibration factor during the field measurements was  $C_1=(2.0\pm 0.4)\times 10^{-8}$  count  $\text{s}^{-1}$  atom $^{-1}$   $\text{cm}^3$ .

### 3.3 Uncertainties and limit of detection

A breakdown of the uncertainties associated with both the  $\text{I}_2$  and I atom sensitivity is shown in Table 1. The limit of detection for measurement systems recording small numbers of discrete events (such as photon counting of fluorescence measurements) may be calculated using Eqs. 11 and 12 (Stevens et al. 1994):

$$\text{LOD} = \frac{S/N}{C_{(\text{I}_2 \text{ or I})}} \sqrt{\left(\frac{1}{m} + \frac{1}{n}\right)} \sigma_b \quad (11)$$

where  $S/N$  is the signal to noise ratio (in this case,  $S/N=1$ ),  $C$  is the pertinent calibration factor (in count  $\text{s}^{-1}$  molecule $^{-1}$ /atom $^{-1}$   $\text{cm}^3$ ) for  $\text{I}_2$  or I accordingly,  $m$  is the number of data acquisition points taken when measuring the  $\text{I}_2$  or I signal and  $n$  is the number of data points taken when measuring the background (usually  $m=n=30$  or  $60$ ) and  $\sigma_b$  is the standard deviation of the background data points given by

$$\sigma_b = \sqrt{\frac{1}{t} S_b} \quad (12)$$

where  $t$  is the time period for the acquisition of a single data point in seconds (usually 1 s) and  $S_b$  is the background signal (count  $\text{s}^{-1}$ ).

For  $\text{I}_2$ , the limit of detection was typically around 1.3 ppb (using  $C_{\text{I}_2}=8\times 10^{-11}$  count  $\text{s}^{-1}$  molecule $^{-1}$   $\text{cm}^3$ ,  $S/N=1$ ,  $m=n=60$ ,  $t=1$  and a typical value for  $S_b$  of 200 count  $\text{s}^{-1}$ ). For iodine atoms, the limit of detection was typically 5.3 ppt (using  $C_1=2\times 10^{-8}$  count  $\text{s}^{-1}$  molecule $^{-1}$   $\text{cm}^3$ , and similar values of  $S/N$ ,  $m$ ,  $n$ ,  $t$  and  $S_b$ ). The limit of detection can be decreased if  $m$  and  $n$  are increased, but there will be a trade off between a lower limit of detection and a low temporal resolution.

**Table 1** Breakdown of the uncertainties associated with both the  $\text{I}_2$  and I atom sensitivity

Parameter	$\pm 1\sigma$ uncertainty (%)
$[\text{O}_3]$	1
$[\text{N}_2\text{O}]$	1
$Ft$	3
$[\text{I}_2]$	11
$C_{\text{I}_2}$	10
$C_1$	10

## 4 Modes of operation

The instrument can be operated in two different modes; either to measure ambient iodine atoms, or to measure the total photolabile iodine loading, through broadband visible photolysis of ambient air, with detection of the iodine atoms released. Direct detection of  $I_2$  at larger concentrations (several orders of magnitude larger than ambient levels), for example, as emitted directly from seaweeds in an enclosed chamber (see Section 5.1), as well as indirect detection by photolysis with a broadband light source and subsequent detection of the iodine atoms formed, may also be accomplished.

### 4.1 Measurement of ambient iodine atoms

When run in ambient sampling mode, air is drawn into the fluorescence cell via the pinhole, and iodine atoms detected as described earlier. During the field deployment, it was found necessary to attach a short length (approximately 1 cm) of blackened Teflon tube over the pinhole in order to block solar radiation from entering the cell, which would otherwise obscure the fluorescence signal (since the CPM is not blind to the extreme lower wavelengths of solar radiation in the troposphere). The Teflon tubing was sealed onto the pinhole with a small piece of neoprene. Subsequent tests showed that this sampling tube did not alter the sensitivity of the instrument to iodine atoms within the uncertainty of the calibration.

Contributions to the signal detected at the CPM arise not only from fluorescence of iodine atoms present in the cell, but also from any dark current on the CPM, scattered light from the cell walls and from aerosol, and potential chemical interferences from other species, including  $I_2$ . In order to obtain the signal from fluorescence of iodine atoms only, this background signal must be subtracted from the total signal recorded, and can only be measured once the iodine atoms have been removed from the sampled air. Ambient air is alternately sampled either directly into the detection cell or is first passed through a darkened pre-reactor, within which iodine atoms are removed through their reaction with ambient ozone:



The darkened pre-reactor was fabricated from a blacked-out Pyrex tube (diameter 50 mm; length 70 cm) with a 90° curve midway, blocked at one end and with a hole to sample via a short length of 6 mm tubing into the fluorescence cell. The lifetime of I atoms with respect to reaction with ambient  $O_3$  is approximately 1.1 s, calculated using  $k_{(12)} = 1.2 \times 10^{-12} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  at 288 K (Atkinson et al. 2007) and using the average  $O_3$  of 30.2 ppb observed during the field deployment. The residence time in the darkened reactor was calculated to be approximately 14 s, in which period almost all (>99.99%) the iodine atoms will be removed through reaction with ambient  $O_3$ . Sampling between the ambient sampling tube and the darkened reactor was modulated manually every 60 s. The modulation between direct sampling and darkened pre-reactor therefore allows subtraction of any contribution to the RF signal from other fluorescing species (including  $I_2$ ) and aerosol scatter.

### 4.2 Measurement of photolabile iodine species

The alternative configuration of the instrument is to measure the total concentration of photolabile iodine-containing species, by employing a broadband visible halogen lamp

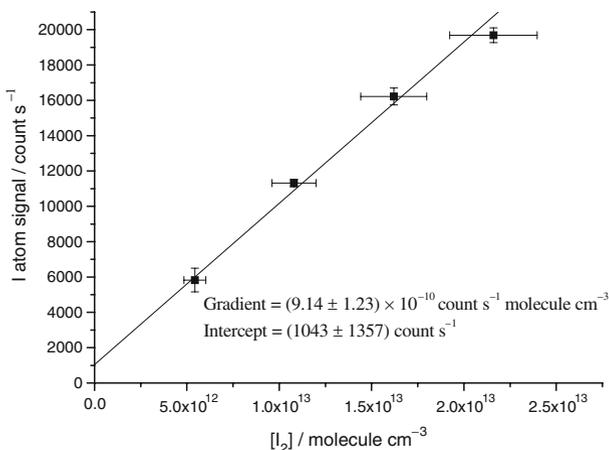
to photolyse ambient air, followed by detection of any iodine atoms so formed. In this configuration, air was passed through the calibration tube, within which photolysis took place, before entering the RF cell. Ambient air was sampled through 6 mm OD PTFE tubing into the calibration flow tube which was mounted over the pinhole in the usual manner. A small pump was attached via a mass flow controller to the excess gas lines to draw air through the PTFE tubing and flow tube at a total flow rate of 7 slm (comprising 5.4 slm through the pinhole and 1.6 slm through the excess line). The broadband lamp was mounted at the bottom of the flow tube, approximately 3 cm above the pinhole and detection cell. The lamp was modulated on/off (details below), and any iodine atoms released from photolysis of iodine-containing species in the ambient air were detected in the usual manner after sampling through the pinhole into the fluorescence cell, with the background signal measured when the photolysis lamp was off. The photolysis lamp also significantly heated the flow tube, raising concern over the potential for thermal dissociation of iodine-containing particles (e.g. nascent  $I_xO_y$  aerosol). To avoid complications from this effect, the temperature of the tube was monitored with a thermocouple, and the lamp was switched off when the temperature reached 40°C (usually after about 5 s). The tube was then allowed to cool to below 27°C before the next photolysis period, a period of 20–30 s. The background signal was taken as an average of 25 data points when the lamp was off and the temperature was between 20°C and 35°C; the measurement signal was taken as the average of 5 data points when the lamp was on and the temperature was between 20°C and 40°C. We do not anticipate any significant thermolysis of iodine-containing species over this temperature range, in the <0.2 s transit time from the warmed region to the RF cell entrance—considering the most likely candidate for thermal decomposition, iodine nitrate, only 1% would decompose at 40°C in this time (decomposition rate from Allan and Plane 2002)—although very little is known concerning the thermal stability of higher iodine oxides.

The total photolabile iodine loading may be represented qualitatively as the iodine atoms released after photolysis of all iodine-containing species, with a relative sensitivity to different species given by the products of their absorption cross sections, I atom photolysis quantum yields, and the emission spectrum of the photolysis lamp. It is likely, at least at the Mace Head site of the field deployment, that the major contributor to iodine atoms from broadband visible photolysis lamp would be  $I_2$ , since  $I_2$  has the largest absorption cross section of the known inorganic iodine species, in the visible wavelength region (Saiz-Lopez et al. 2004), and comparison of previous observations of  $I_2$  and iodocarbons at Mace Head (Saiz-Lopez and Plane 2004; Carpenter et al. 1999) indicate that level of molecular iodine is much greater (by nearly two orders of magnitude) than the sum of the measured organic iodine species. The sensitivity of the total photolabile iodine measurement to molecular iodine was obtained by measuring the iodine atom signal resulting from photolysis of a known concentration of  $I_2$  in air, using the broadband photolysis lamp:

$$C_{PL,I_2} = \frac{S_{PL}}{[I_2]} \quad (14)$$

where  $C_{PL,I_2}$  is the calibration factor for the photolysis lamp,  $S_{PL}$  is the iodine atom signal measured with the photolysis lamp turned on (after subtraction of background, including  $I_2$  fluorescence) in the presence of a known concentration of  $I_2$ . Figure 7 shows the variation in  $S_{PL}$  with  $[I_2]$ ; the gradient of this graph determines the sensitivity of the

**Fig. 7** Relationship between iodine atom signal and  $I_2$  concentration, in the photolabile configuration ( $I_2$  photolysis within the calibration tube, by the broadband photolysis lamp); the gradient of the line determines  $C_{I_2,PL}$  (eq. 14).  $y$ -Axis error bars represent the standard deviation of the fluorescence signal;  $x$ -axis error bars represent the uncertainty in the  $I_2$  concentration. The linear regression takes into account both  $x$ - and  $y$ -errors



photolabile iodine measurement (to molecular iodine),  $C_{PL,I_2} = (9.1 \pm 1.2) \times 10^{-10}$  counts  $s^{-1}$  molecule $^{-1}$   $cm^3$ . Note that  $C_{PL,I_2}$  implicitly incorporates the quantum yield for I atom production from  $I_2$  photolysis. Equation 14 may then be applied to the measured iodine signal from ambient air, in the photolabile measurement configuration, to yield the concentration of  $I_2$  present, under the assumption that  $I_2$  is the species which dominates this signal.

In a laboratory setting, molecular iodine can also be detected through its direct fluorescence without the need for prior photolytic conversion to iodine atoms, but the sensitivity ( $C_{I_2}$ ,  $8 \times 10^{-11}$  counts  $s^{-1}$  molecule $^{-1}$   $cm^3$ ) is approximately an order of magnitude worse than that obtained through I atom detection following photolysis, insufficient for atmospheric measurements. Comparison of the sensitivity of the photolabile measurement with that for iodine atoms yields the photolysis efficiency for  $I_2$ :

$$\text{Photolysis efficiency} = \frac{C_{PL,I_2}}{2 \times C_I} \quad (15)$$

where the factor of 2 accounts for the iodine atom quantum yield. Application of Eq. 15 to the relevant calibration factors determines an  $I_2$  photolysis efficiency of 2.3%.

## 5 Mace head field measurements

The instrument was deployed for the first time in the field in August 2007 at the Mace Head Atmospheric Research Station on the west coast of Ireland (53° N, 9° W). This location is well known for boundary layer halogen activity (e.g. Alicke et al. 1999; Carpenter et al. 2000; Allan et al. 2000); a description of the site and prevailing chemical and meteorological conditions during summer is given by Heard et al. (2006). Measurements in the iodine atom configuration were performed just to the north of the shore-side laboratory building, at a position with open access to airflow from the ocean, at an elevation of approximately 5 m above mean sea level. In the photolabile iodine configuration, the instrument was located within the shoreline laboratories, and ambient air was sampled on the western (ocean) side from approximately 2 m above ground level.

### 5.1 Emission of molecular iodine from seaweed

Whilst on site at Mace Head, the instrument was used to observe direct emission of  $I_2$  from seaweed. Small amounts of six different species of seaweed, including *Himanthalia elongata*, *Palmaria palmata*, and *Laminaria digitata* were gathered from rock pools exposed during one of the lowest tides.

Synthetic air was flowed through a large Pyrex flask (volume=2 l) and into the calibration tube at a rate of 10 slm. After the background signal had been recorded, 50 g of the (wet) seaweed sample was added to the Pyrex vessel. The broadband visible lamp was used to photolyse any  $I_2$  (and any other photolabile species present) in the calibration tube for about 5 seconds at approximately one minute intervals. Only one type of seaweed was observed to emit detectable levels of  $I_2$ : *Laminaria digitata*.

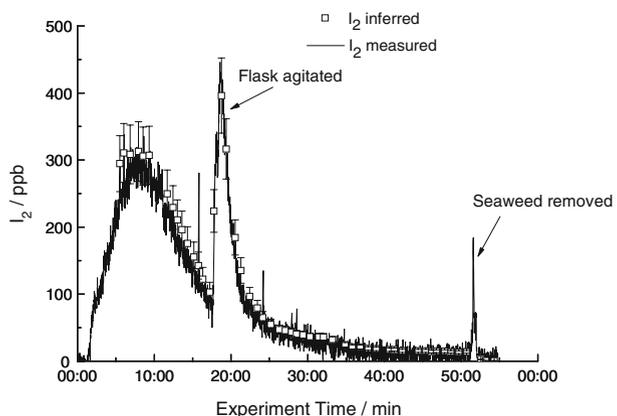
As shown in Fig. 8, 50 g of *Laminaria digitata* was observed to emit  $I_2$  at levels of up to 300 ppb, which decreased over the period of about an hour. Agitation of the seaweed in the vessel released a further burst of  $I_2$ . These high levels were observed without intentionally stressing the seaweed; no  $O_3$  was present since the bath gas used was (cylinder) compressed air, and the actinic intensity (indoors, within the Pyrex vessel) was moderate. The seaweed was kept in seawater until tested (within 30 minutes of harvesting), and was not dried before testing, although the sample gradually dried out over time due to the flow through the vessel. The leaves of the seaweed were not deliberately damaged or cut. Palmer et al. (2005) suggested that under oxidative stress (caused by either exogenous hydrogen peroxide, gaseous ozone or a solution of oligogulonates, known elicitors of oxidative stress) the seaweed *Laminaria digitata* increased production of  $I_2$ . Therefore, levels of  $I_2$  emitted from this type of seaweed under atmospheric conditions could be significantly greater than those reported here.

In the seaweed experiment,  $I_2$  concentrations were obtained both indirectly, via photolysis and subsequent detection of the iodine atoms released, and by directly observing the (much weaker)  $I_2$  fluorescence. The excellent agreement between the two determinations of  $[I_2]$  apparent from the plot (Fig. 8) gives confidence in the calibration methods used.

### 5.2 Measurement of ambient iodine atoms

Iodine atoms were observed during the day of the 29 August 2007. For these measurements, the instrument was located about 45 m from the high-tide water line, and

**Fig. 8** Time series showing levels of  $I_2$  emitted directly from a 10 slm flow of synthetic air over a 50 g sample of *Laminaria digitata* seaweed within a 2 l flask. The black line represents  $[I_2]$  measured directly. The open squares represent  $[I_2]$  inferred after photolysis with the broadband visible lamp and detection of the iodine atoms formed; the y-axis errors are given by the uncertainty in the gradient of Fig. 7



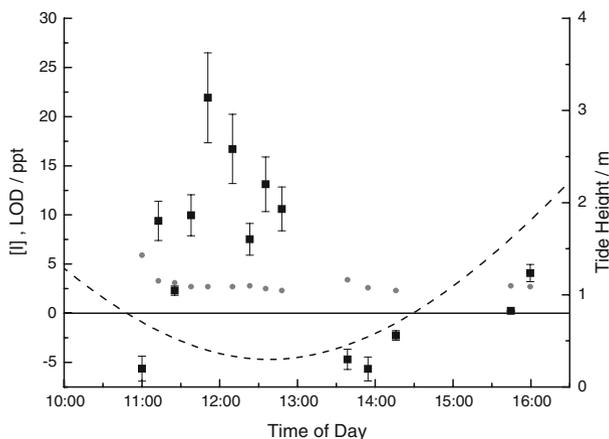
approximately 100 m from the low-tide line. Figure 9 presents a plot showing the time series for iodine atoms; the measurements are well above the limit of detection (calculated from the measured calibration factor and the background signal) of the instrument, which was around 3 ppt for a 300 s integration period. The peak in iodine atoms correlates strongly with the local low tide (data recorded at Roundstone Bay; Admiralty Easytide database; August 2007), which supports the hypothesis of a local coastal source of  $I_2$  and/or the alkyl iodide precursors. It is interesting to note that the rock pools containing *Laminaria digitata*, which we have shown directly emits  $I_2$ , and other seaweeds of a similar type (e.g. *Laminaria saccharina*) were only exposed at the very lowest tides experienced at Mace Head, as was the case on 29 August. On inspection of the data recorded at a temporal resolution of 1 s, it was apparent that the iodine atom concentrations are highly variable, and fluctuate greatly on this timescale (relative standard deviation of 300%), implying a highly variable atmospheric iodine flux. Given the close proximity to the exposed pools containing seaweed known to emit  $I_2$  (approximately 50–100 m), and the short lifetime of molecular iodine with respect to photolysis (5–10 s when the sun is overhead; Saiz-Lopez et al. 2004), any variation in the wind speed and direction will greatly affect the processing time and hence the concentration of atomic iodine.

The concentrations reported here may be overestimates of the levels of atomic iodine sampled for two reasons: If a significant proportion of aerosol particles were lost to the walls of the darkened reactor, then the background signal measurement might appear lower than during the ambient measurement period, when the increased aerosol would add to the light scattering and increase the apparent fluorescence signal. Secondly, if significant levels of iodine atoms were lost during calibration, as a consequence of wall encounters during turbulence introduced by the  $O_2$  addition, the calibration constant ( $C_1$ ) would be reduced, and the retrieved ambient iodine atom levels overestimated, although we believe this to be a minor effect. Developments are in progress to address these issues (improving the various flow geometries). The values presented may therefore represent upper limits on the levels of atomic iodine present.

### 5.3 Measurement of photolabile species

Experiments to measure the total photolabile iodine content of ambient air were carried out during several night-time and day-time periods throughout the Mace Head campaign. At

**Fig. 9** Time series (local time IST) showing ambient I atoms (5 min integration period) at Mace Head on 29 Aug 2007. The grey circles represent the limit of detection for a 5 min integration period. The dashed line represents the tide height

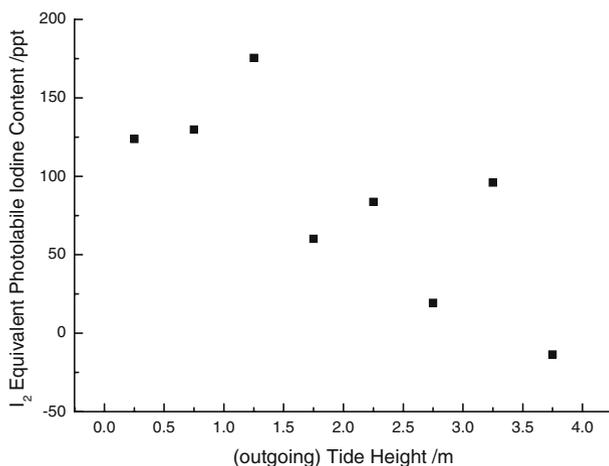


night-time, in particular, it is likely that the majority of the photolabile iodine will be released from molecular iodine, and levels of  $I_2$  will build up in the absence of solar photolysis. Figure 10 shows a plot of inferred  $I_2$  mixing ratio (calculated using Eq. 14) as a function of tide height during the night of the 16/17 August 2007. It is apparent that the levels of  $I_2$  are higher at low tide, which is in agreement with the hypothesis that seaweed species such as *Laminaria digitata* emit high levels of  $I_2$  directly when exposed—an equivalent plot of measured  $I_2$  as a function of tidal height using observations made during the NAMBLEX campaign by Saiz-Lopez et al. (2006) shows a strikingly similar trend over the same tidal height range.

#### 5.4 Comparison with previous observations

Gas-phase reactive inorganic iodine species (IO,  $I_2$ ) have been observed at Mace Head previously using long-path differential optical absorption spectroscopy (LP-DOAS) techniques, which determine average mixing levels along a 4–8 km lightpath (Alicke et al. 1999; Allan et al. 2000; Saiz-Lopez and Plane 2004), while in situ measurements of  $I_2$  have been performed at Mace Head using cavity ring-down spectroscopy (CRDS; Bitter et al. 2005), and at nearby locations using denuder tubes coupled with ICP/MS analysis (Saiz-Lopez et al. 2006). The LP-DOAS observations of IO have ranged from <1 to 7 ppt, with  $I_2$  observed at up to 93 ppt (night) and 20 ppt (day). The in situ observations have detected  $I_2$  at up to 94 ppt (CRDS) and 115 ppt (denuder samples), supporting the conclusion from the observations reported here, that local littoral sources dominate the inorganic iodine activity at Mace Head. Saiz-Lopez et al. (2006) have modelled the inorganic iodine chemistry anticipated for different iodine source distributions, ranging from an even distribution along the DOAS beampath through to solely local shoreline emissions, constraining their model scenarios with the LP-DOAS, CRDS and denuder observations. In the shoreline sources only case, they predict daytime levels of  $I_2$ , IO and I atoms of 80–100, 40–50 and 15–20 ppt respectively, in good qualitative agreement with the observations reported here. Most recently, Commane et al. (2008) have reported new in situ measurements of IO radicals by laser-induced fluorescence, performed at Mace Head during August 2008; the observations (up to 34 ppt IO, correlated with tide height) are consistent with the I atom and photolabile

**Fig. 10** Photoproducted iodine atom signal, depicted as  $I_2$  mixing-ratio equivalent, after photolysis of ambient air, as a function of tide height at Mace Head during the night of 16/17 Aug 2007 (20:30–03:00 local time)



iodine measurements reported in this work, and with the overall inorganic iodine distribution simulated by Saiz-Lopez et al. (2006), for the shoreline sources only case.

## 6 Summary

A portable field instrument employing resonance fluorescence has been developed and has been shown to be sufficiently sensitive for the selective measurement of ambient levels of atomic iodine. The limit of detection achieved with this instrument, 5.3 ppt, is significantly lower than has been reached previously in systems for laboratory-based kinetics/photochemistry experiments. Observations made at the Mace Head Atmospheric Research Station in August 2007 indicate that iodine atoms are present at levels up to  $22 \pm 5$  ppt at low tide. This measurement represents the first direct observation of iodine atoms in the marine boundary layer. The instrument has also been used to measure the total photolabile iodine loading of ambient air, at Mace Head probably corresponding predominantly to  $I_2$ , and hence to infer the  $I_2$  abundance; observations overnight determined molecular iodine levels of up to 200 ppt. Future modifications to the instrument will improve the sensitivity of the photolabile detection technique, including the addition of a selective photolysis capability. With some further development, the instrument could be used in a variety of measurement modes, for example on ship or aircraft platforms.

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